CORRELATION OF DIOXIN ANALOGUES CONCENTRATIONS BETWEEN AMBIENT AIR AND PINE NEEDLE IN JAPAN 1

Komichi Ikeda¹, Teiichi Aoyama², Atsushi Takatori³, Hideaki Miyata⁴ and Patrick Pond⁵

Environmental Research Institute Inc., Tokyo Japan¹⁻³, Food Hygiene Laboratory, Faculty of Pharmaceutical Sciences, Setsunan University, Osaka Japan⁴, Environmental Science Division, Maxxam Analytics Inc., Ontario Canada⁵

Introduction

The needle of Japanese black pine (hereafter abbreviated as pine needles) has been used as an

effective bio-monitor for ambient air. Miyata et al. of Setsunan University has reported that the pine needles can accumulate dioxin analogues through photosynthesis and respiration.¹ Fig.1 shows the result of the accumulation of PCDDs and PCDFs (D/F) reported by Miyata et al., where sampling took place every 2 weeks at Hirakata City, Osaka¹. The D/F in the ambient air is absorbed quite rapidly into the pine needle at the early stages. However, after 4 months the accumulation reaches a steady state in accordance to the pollution levels of the local ambient air conditions.

The purpose of this study is to make clear the correlation of average D/F concentration between pine needles and ambient air. If there is a clear correlation between the two matrices, it is possible to estimate the yearly average D/F concentration in ambient air by analyzing pine needle sampled more than 6 months after the pine needle sprout.

In order to prove this hypothesis, we analyzed pine needles from the neighboring area of long term continuous ambient air monitoring points at U.S. Naval Air Facility Atsugi, Kanagawa prefecture.²

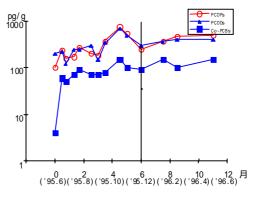
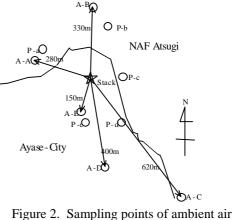


Figure 1. Time course of accumulation of D/F in pine needles



and pine needle

Method

The sampling of pine needles was done from November to December of 1999. Our target pine needle of 2 years old was sampled at ca. 1.5m height above the ground level. The sampling was ca. 100g per each sample points. The analytical methodology for the determination of D/F is indicated in Fig. 3. The details are described elsewhere.¹

 Sample (50 g) Cut into a size of 3 cm Dried by lyophilization Lyophilized sample Macerated with an Ultra-Turrax using toluene (500ml) Extraction with toluene under reflux (4hr) Addition of silica gel (50g) and leave it a whole day and night Evaporated to dryness and dissolved with n-hexane (10ml) 30% aliquot of extract Addition of ¹³C₁₂-PCDDs and PCDFs internal standards Multi layer silica gel column chromatography Eluted with n-hexane (210 ml)
 Dried by lyophilization 2. Lyophilized sample Macerated with an Ultra-Turrax using toluene (500ml) Extraction with toluene under reflux (4hr) Addition of silica gel (50g) and leave it a whole day and night Evaporated to dryness and dissolved with n-hexane (10ml) 3. 30% aliquot of extract Addition of ¹³C₁₂-PCDDs and PCDFs internal standards Multi layer silica gel column chromatography
 2. Lyophilized sample Macerated with an Ultra-Turrax using toluene (500ml) Extraction with toluene under reflux (4hr) Addition of silica gel (50g) and leave it a whole day and night Evaporated to dryness and dissolved with n-hexane (10ml) 3. 30% aliquot of extract Addition of ¹³C₁₂-PCDDs and PCDFs internal standards Multi layer silica gel column chromatography
 Macerated with an Ultra-Turrax using toluene (500ml) Extraction with toluene under reflux (4hr) Addition of silica gel (50g) and leave it a whole day and night Evaporated to dryness and dissolved with n-hexane (10ml) 3. 30% aliquot of extract Addition of ¹³C₁₂-PCDDs and PCDFs internal standards Multi layer silica gel column chromatography
 Extraction with toluene under reflux (4hr) Addition of silica gel (50g) and leave it a whole day and night Evaporated to dryness and dissolved with n-hexane (10ml) 3. 30% aliquot of extract Addition of ¹³C₁₂-PCDDs and PCDFs internal standards Multi layer silica gel column chromatography
 Addition of silica gel (50g) and leave it a whole day and night Evaporated to dryness and dissolved with n-hexane (10ml) 3. 30% aliquot of extract Addition of ¹³C₁₂-PCDDs and PCDFs internal standards Multi layer silica gel column chromatography
Evaporated to dryness and dissolved with n-hexane (10ml) 3. 30% aliquot of extract Addition of ¹³ C ₁₂ -PCDDs and PCDFs internal standards Multi layer silica gel column chromatography
 30% aliquot of extract Addition of ¹³C₁₂-PCDDs and PCDFs internal standards Multi layer silica gel column chromatography
Addition of ¹³ C ₁₂ -PCDDs and PCDFs internal standards Multi layer silica gel column chromatography
Multi layer silica gel column chromatography
Eluted with n-hexane (210 ml)
Concentrated to 10 ml in n-hexane
Alumina column chromatography (neutral, activity I)
Separated into 1 st and 2 nd fractions
Concentrated to 20 µl in n-decane
4. 2 nd fraction
HRGC-HRMS-SIM analysis (R=7000 – 10000)

Figure 3. Outline of analytical method for PCDDs and PCDFs in black pine needle¹

Results and Discussion

Table 1 shows the D/F TEQ concentration in pine needles in relation to the distance and direction from an industrial waste incineration facility on site. The concentration of the background point was 2.4 pg-TEQ/g dry weight. The levels of Site P-b (North of the incinerator) and P-e (South of the incinerator) as a target point were respectively 53.1 pg-TEQ/g and 30.6 pg-TEQ/g. On the contrary, Sites P-a, P-c and P-d were 4.1, 7.7 and 11 pg-TEQ/g, respectively, although the distance from the incinerator at these 3 sites were nearly equal to those of Site P-b and P-e. This shows that the D/F concentration in pine needles will be affected not only by the distance from the incinerator, but also by the geographical and yearly meteorological conditions of the local area. Figure 4 shows the estimated spatial D/F concentration contour on the basis of D/F in pine needles analyzed by the Spline interpolation method⁴.

In addition, Table 2 shows the ambient D/F concentration in approximately the same area of the pine sampling points. Figure 5 shows the actual monitoring data of D/F TEQ concentration in ambient air for 56 continuous days.²

-	, It		
Sampling	Distance	Direction	Pine
Point			Needle
Site P-a	280m	NW	4.1
Site P-b	250	Ν	53.1
Site P-c	180	Е	11.0
Site P-d	200	SE	7.7
Site P-e	170	S	30.6
Back	1500	NNE	2.4
Ground			

Table 1. D/F concentration in pine needle at NAF Atsugi. (Unit: pg-TEQ/g dry weight)

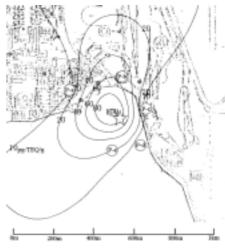


Figure 4. Spatial D/F concentration of pines analyzed by Spline interpolation method

Table 2 Comparison of D/F concentration between ambient air and pine needle

Air	Min.	Max	Ave.	Period	Pine	An. Ave.	Ratio	
A-A	0.085	3.3	0.59	Summer	P-a	4.1		
A-B	0.097	53	7.4	Summer	P-b	53.1		Source: Japan-US Joint
A-C	0.031	1.5	0.28	Summer	P-c	11.0		monitoring survey in NAF Atsugi ² survey of ambient air dioxins in
A-D	0.062	1.3	0.50	Winter	P-d	7.7		
A-E	0.11	21	1.4	Winter	P-e	30.6		
	Ave.		2.0(A)			21.3(B)	10.7	Seya ³

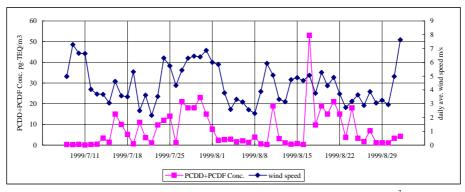


Figure 5 Ambient air D/F concentration at Site B. unit:pg-TEQ/m³ Source: Japan-US Joint monitoring survey in NAF Atsugi²

As shown in Table 2, the ratio of D/F concentration in pine needles and ambient air was calculated to reveal the correlation between the two different matrices. The ratio was 1:10.7, the units were pg-TEQ/g for pine needle and pg-TEQ/m³ for ambient air. Since the D/F concentration in pine needles is estimated as a long term average level, and the ambient air concentration is the average of 56 days continuous monitoring, it is necessary to unify both data in a long term average concentration to clearly identify the ratio between the two matrices. Therefore, three dimensional aero-dynamic air dispersion simulation model was used. The geographical and meteorological conditions were applied to estimate the yearly average of ambient air D/F concentration at the pine needle sampling points.⁵ Consequently, the concentration ratio of pine needle versus ambient air was estimated to be 10.7, which was derived from the actual monitored data shown in Table 2.

	Ambient Air	Pine Needle	Pine / Air	Unit: Air pg-TEQ/m ³
Site P-a	0.60	4.1	6.8	Pine pg-TEQ/g
Site P-b	4.75	53.1	11.2	
Site P-c	1.19	11.0	9.2	
Site P-d	0.80	7.7	9.6	
Site P-e	2.40	30.6	12.8	
Ave.	1.95	21.3	10.9	

Table 3 Correlation between Ambient air and Pines in Annual average.

Acknowledgment

This project could not be possible without the cooperation of Shozo Kajiyama of Mirai Shimin Law Office and the Naval Air Facility of Atsugi. Authors also express hearty thanks to those members of Setsunan University and the staff of Maxxam Analytics Inc. for analyzing the pine needles.

References

1.Masaru Ikeda, Hideaki Miyata, (1997) Time Trend on Accumulation of PCDDs, PCDFs and Co-PCBs in Young Pine Needle (Thesis), Setsunan Univ.

2. Japan-Us Joint Monitoring Survey of Ambient Air Dioxins in NAF Atsugi, (1999) Japan Environment Agency

3. Survey of Ambient Air Dioxins in Seya, Dec.1999 to Feb.2000, (2000) Kanagawa Prefecture, Japan

4. Yukio Onishi, (1975) 2 Dimensional Interpolation by Using Spline Method, Japan Oceanographic Society, Japan 31

5.Teiichi Aoyama, Atsushi Takatori, Estimation of the Dioxins Emission From the Enviro-Tech Incineration Stacks at NAF Atsugi, (2000) Earth Tech. Inc., U.S. Department of Navy, Pacific Division.